NucleoSpin® Plasmid (NoLid)

1 Kit contents

	NucleoSpin® Plasmid (NoLid)	
REF	**12 preps 740499.12C**	**240 preps 740499.240C**
Resuspension Buffer A1	15 mL (add RNase A before first use)	100 mL (add RNase A before first use)
Lysis Buffer A2	15 mL	100 mL
Neutralization Buffer A3	20 mL	200 mL
Wash Buffer AW	30 mL	2 x 100 mL
Wash Buffer A4 (Concentrate)	6 mL (add 24 mL ethanol before first use)	100 mL (add 400 mL ethanol before first use)
Elution Buffer AE	30 mL	125 mL
RNase A (lyophilized)	6 mg	40 mg
NucleoSpin® Plasmid (NoLid) Columns (white rings)	12	4 x 60
User manual	1	1

2 How to use the kit

Please see the protocol information how to use the kit (see next pages). For further questions and more detailed information, please contact MACHEREY-NAGEL at tech-bio@mn-net.com for protocol information how to use the kit on specific robotic instruments.

For storage conditions, product use restrictions, and safety information, please see the general NucleoSpin® Plasmid (NoLid) user manual.

NucleoSpin® kits on QIAcube®
MN is not recommending to use this kit on specific robots. The use of NucleoSpin® kits on theQIAcube® is solely at your own discretion. MACHEREY-NAGEL is not responsible for loss of warranty claims or other consequences.

3 General information

Application:	Plasmid DNA
Kit:	NucleoSpin® Plasmid (NoLid) (REF 740499.240C) instead of: QIAprep® Spin Miniprep Kit
Sample material:	Up to 5 mL LB cultures (for TB or 2x YT medium use up to 2.5 mL culture volume)
Protocol name:	Plasmid DNA purification using the QIAprep® Spin Miniprep Kit (with PB wash) DNA_QIAprepMiniprep_UpTo5mlLBCultures_Standard_V3
Editable parameters:	Elution volume: 30–100 µL; default 50 µL

4 Using the kit

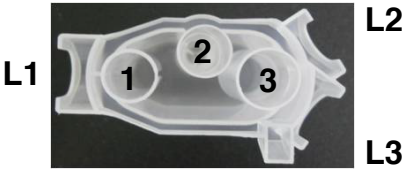
1. Fill the designated buffer bottles with the buffers according to the buffer table below (make sure that RNase A was added to Buffer A1 in advance).
2. Harvest bacterial cells of up to 5 mL LB cultures by centrifugation at 6800 x *g* for 3 minutes.
3. Place Sample Tubes (2 mL Safe-Lock microcentrifuge tubes) containing the pelleted bacterial cells into sample rack (shaker).
4. Insert disposable Filter Tips 1000 µL and 1000 µL wide-bore.
5 General equipment setup is shown below.

5 Additional materials

Refer to the QIAcube® protocol sheet for required consumables (e.g., sample tubes, collection tubes, instrument accessories, disposable tips, etc.) and software requirements.

6 Rotor adapter

Position	Labware	Lid position
1	NucleoSpin® Plasmid (NoLid) Column	L1
2	–	–
3	1.5 mL collection tube*	L3



* Sarstedt, Micro tube 1.5 mL Safety Cap

7 Buffers (Reagent Bottle Rack)

Position	MN Reagent	Replaced QIAGEN® Reagent
1	Buffer A1	Buffer P1
2	Buffer A2	Buffer P2
3	Buffer A3	Buffer N3
4	Buffer AW	Buffer PB
5	Buffer A4	Buffer PE
6	Buffer AE	Buffer EB

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